

The MAPK model

Tapesh Santra, Walter Kolch, Boris N. Kholodenko

...

ODE simulation: The ODE model for our simulation experiment is shown in Tab.1.

Table 1: MAPK model

Receptor module		
EGF binds to EGFR	$R + L = RL$	$ka = 0.001, kd = 0.01$
EGF-EGFR complex dimerization	$RL + RL = RD$	$ka = 0.001, kd = 0.01$
EGF-EGFR complex phosphorylation	$RD \rightarrow pRD$	$k = 0.01$
EGFR complex dephosphorylation	$pRD \rightarrow RD$	$\frac{Vm_1}{(Km_1 + pRD)}$; $Vm_1 = 1, Km_1 = 10$
Adapter module		
Phosphorylation of Shc by EGFR	$Shc \rightarrow pShc; pRD$	$\frac{kcat_1 \times pRD \times Shc}{(K_1 + Shc)}$; $kcat_1 = 0.1, K_1 = 5$
Dephosphorylation of Shc	$pShc \rightarrow Shc$	$\frac{Vm_2 \times pShc}{(Km_2 + pShc)}$; $Vm_2 = 0.08, Km_2 = 200$
Shc binds to Grb2SOS complex	$pShc + GS = pShc-GS$	$ka = 0.001, kd = 0.01$
ppERK phosphorylates and inhibits GS	$GS \rightarrow iGS; ppERK$	$\frac{kcat_9 \times GS \times ppERK}{(K_9 + GS)}$; $kcat_9 = 0.01, K_9 = 10$
iGS dephosphorylation	$iGS \rightarrow GS$	$\frac{Vm_{10} iGS}{(Km_{71} + iGS)}$; $Vm_{10} = 0.05, Km_{71} = 10$
Initiator module		
Phosphorylation of RasGDP by Shc bound Grb2SOS	$RasGDP \rightarrow RasGTP; pShc-GS$	$\frac{kcat_2 \times pShc \times GS \times RasGDP}{(K_2 + RasGDP)}$; $kcat_2 = 0.001, K_2 = 10$
Dephosphorylation of RasGTP	$RasGTP \rightarrow RasGDP$	$\frac{Vm_3 RasGTP}{(Km_3 + RasGTP)}$; $Vm_3 = 0.1, Km_3 = 5$
MAP3K module		
Raf activation stage 1	$Raf \rightarrow aRaf; RasGTP, ppERK$	$\frac{kcat_3 \times RasGTP \times Raf}{((K_{31} + Raf + aRaf \frac{K_{31}}{K_{32}})(1 + \frac{ppERK}{K_i}))}$; $kcat_3 = 0.1, K_{31} = 300, K_{32} = 20; K_i = 5$
Raf activation stage 2	$aRaf \rightarrow aaRaf; RasGTP, ppERK$	$\frac{kcat_4 \times RasGTP \times aRaf}{((K_{31} + Raf + aRaf \frac{K_{31}}{K_{32}})(1 + \frac{ppERK}{K_i}))}$; $kcat_4 = 0.1, K_{31} = 300, K_{32} = 20; K_i = 5$
Raf deactivation stage 1	$aaRaf \rightarrow aRaf;$	$\frac{Vm_4 \times aaRaf}{(Km_{41} + aaRaf + (aRaf \frac{Km_{41}}{Km_{42}}) + Raf \frac{Km_{41}}{Km_{43}})}$; $Vm_4 = 0.05, Km_{41} = 22, Km_{42} = 18; Km_{43} = 80$
Raf deactivation stage 1	$aRaf \rightarrow Raf;$	$\frac{Vm_5 \times aRaf}{(Km_{41} + aaRaf + (aRaf \frac{Km_{41}}{Km_{42}}) + Raf \frac{Km_{41}}{Km_{43}})}$; $Vm_5 = 0.05, Km_{41} = 22, Km_{42} = 18; Km_{43} = 80$
MAP2K module		
MEK phosphorylation	$MEK \rightarrow pMEK; aaRaf$	$\frac{kcat_5 \times MEK \times aaRaf}{(K_{41} + MEK + pMEK \frac{K_{41}}{K_{42}})}$; $kcat_5 = 0.1, K_{41} = 300; K_{42} = 20$
MEK doublephosphorylation	$pMEK \rightarrow ppMEK; aaRaf$	$\frac{kcat_6 \times pMEK \times aaRaf}{(K_{41} + MEK + pMEK \frac{K_{41}}{K_{42}})}$; $kcat_6 = 0.1, K_{41} = 300; K_{42} = 20$
MEK dephosphorylation stage 1	$ppMEK \rightarrow pMEK; ppERK$	$\frac{Vm_6 \times ppMEK \times (1 + A \frac{ppERK}{K_{mp}})}{((Km_{51} + ppMEK + (pMEK \frac{Km_{51}}{Km_{52}}) + MEK \frac{Km_{51}}{Km_{53}}) * (1 + \frac{ppERK}{K_{mp}}))}$; $Vm_6 = 0.09, Km_{51} = 22, Km_{52} = 18; Km_{53} = 80; K_{mp} = 100$
MEK dephosphorylation stage 2	$pMEK \rightarrow MEK; ppERK$	$\frac{Vm_7 \times pMEK \times (1 + A \frac{ppERK}{K_{mp}})}{((Km_{51} + ppMEK + pMEK \frac{Km_{51}}{Km_{52}} + MEK \frac{Km_{51}}{Km_{53}}) * (1 + \frac{ppERK}{K_{mp}}))}$; $Vm_7 = 0.09, Km_{51} = 22, Km_{52} = 18; Km_{53} = 80; K_{mp} = 100$
MAPK module		
ERK phosphorylation	$ERK \rightarrow pERK; ppMEK$	$\frac{kcat_7 \times ERK \times ppMEK}{(K_{51} + ERK + pERK \frac{K_{51}}{K_{52}})}$; $kcat_7 = 0.1, K_{51} = 300; K_{52} = 20$
ERK doublephosphorylation	$pERK \rightarrow ppERK; ppMEK$	$\frac{kcat_8 \times pERK \times ppMEK}{(K_{51} + ERK + pERK \frac{K_{51}}{K_{52}})}$; $kcat_8 = 0.1, K_{51} = 300; K_{52} = 20$
ERK dephosphorylation stage 1	$ppERK \rightarrow pERK$	$\frac{Vm_8 \times ppERK}{(Km_{61} + ppERK + pERK \frac{Km_{61}}{Km_{62}} + ERK \frac{Km_{61}}{Km_{63}})}$; $Vm_8 = 0.05, Km_{61} = 22, Km_{62} = 18, Km_{63} = 80$
ERK dephosphorylation stage 2	$pERK \rightarrow ERK$	$\frac{Vm_9 \times pERK}{(Km_{61} + ppERK + pERK \frac{Km_{61}}{Km_{62}} + ERK \frac{Km_{61}}{Km_{63}})}$; $Vm_9 = 0.05, Km_{61} = 22, Km_{62} = 18, Km_{63} = 80$

The initial concentrations for the MAPK model are given in Tab.2.

siRNA knockdown experiments siRNA knockdown of a gene results in attenuated level of the corresponding proteins. Therefore we simulated knockdown experiments by reducing the initial concentrations of the corresponding proteins. The initial concentrations regarding the knock down experiments are shown in Tab.3.

Table 2: Initial concentrations for the MAPK model

Initial concentrations	
	L=1
	R=100
	SHC = 300
	GS=100
	RasGDP =100
	RAF_total= Raf + aRaf+ aaRaf= 100
	MEK_total = MEK + pMEK + ppMEK=100
	ERK_total = ERK + pERK + ppERK=100

Table 3: Initial concentrations for the knockdown experiments

Replicate 1: 80% knockdown efficiency						
Module:	Exp. no.					
	1	2	3	4	5	6
R	20	100	100	100	100	20
Shc	300	60	300	300	300	300
RasGDP	100	100	20	100	100	100
RAFtotal	100	100	100	20	100	100
MEKtotal	100	100	100	100	20	100
ERKtotal	100	100	100	100	100	20

Replicate 2: 60% knockdown efficiency						
Module:	Exp. no.					
	1	2	3	4	5	6
R	40	100	100	100	100	20
Shc	300	120	300	300	300	300
RasGDP	100	100	40	100	100	100
RAFtotal	100	100	100	40	100	100
MEKtotal	100	100	100	100	40	100
ERKtotal	100	100	100	100	100	40

Replicate 3: 40% knockdown efficiency						
Module:	Exp. no.					
	1	2	3	4	5	6
R	60	100	100	100	100	20
Shc	300	180	300	300	300	300
RasGDP	100	100	60	100	100	100
RAFtotal	100	100	100	60	100	100
MEKtotal	100	100	100	100	60	100
ERKtotal	100	100	100	100	100	60

SDE simulation: Each ordinary differential equation of the following form

$$\frac{dx}{dt} = V(\mathbf{x}) - D(\mathbf{x}) \quad (1)$$

was converted into the a stochastic differential equation of the following form [1]

$$\frac{dx}{dt} = V(\mathbf{x}) - D(\mathbf{x}) + c_1 \left(\sqrt{V(\mathbf{x})}\eta_v + \sqrt{D(\mathbf{x})}\eta_d \right) \quad (2)$$

Here, η_v and η_d are independent Gaussian white noises and $c_1 = 0.01$ is a multiplicative constant which controls the amplitude of the molecular noise. The scheme was adopted from [1].

References

- [1] T. Schaffter, D. Marbach, and Floreano D. Genenetweaver: In silico benchmark generation and performance profiling of network inference methods. *Bioinformatics*, 26:2263–70, 2011.